Clinicopathological studies on the effect of spirulina in culture Nile tilapia.

Abdalla O.A., Eissa I. A.*, Amina A. Dessuki**, Omnia E. Kilany and Shimaa M.Elbahar***

Department of Clinical Pathology; *Department of Fish disease and Management and **Department of Pathology, Fac. Vet. Med., Suez Canal University, and ***Fac. Vet. Med. Suez Canal University, Egypt.

Abstract

This study was undertaken to study the effect of spirulina (Arthrospir platensis) on some serum biochemical parameters of O.niloticus. A total of 270 fish (50±5 g) were randomly distributed into six groups each at a rate of 15 fish per aquarium and fed on a diet containing 0.0, 5.0 or 10.0 g spirulina/kg diet for 6 weeks. Each subdivided into three equal replicates. After the feeding trial, fish of each treatment were challenged by pathogenic Pseudomonas. fluorescens which was given by I/P injection. The blood samples were taken after 4and 6weeks for serum biochemical examinations. The results showed that spirulina improved serum AST, ALT, total protein, albumin, globulin, glucose, creatinine, uric acid and cholesterol in groups supplemented with spirulina. Moreover, spirulina enhanced histopathological lesions of fish infected with P. fluorescens. These results indicate that spirulina supplementation is promising for disease prevention in tilapia culture, and the optimum level of spirulina in fish diet is 10.0 g per kg diet.

Introduction:

Functional feed additives strategy has recently gained considerable attention. From nutritional point of view; it does not only provide the essential nutrients required for normal physiological functioning, but also serve as a medium by which fish receive other components that may positively affect their health (*Ibrahem et al*, 2010).

Spirulina (*Spirulina platensis*) is a freshwater blue-green filamentous alga, and it is receiving increasing attention for its bioactive

components such as vitamins, protein (60–70%),minerals. polyunsaturated fatty acids. carotenes and other pigments that antioxidants (Madhava et al, 2000; Lin et al, *2007*). Moreover, Bermejo et al reported (2008)that antioxidant capacities of Spirulina protean extract are attributable to the biliproteins contained in this microalga, such as phycocyanin. The previous authors suggested that spirulina could be used to produce a dietary natural antioxidant supplement or added to healthy

food products, such as cereals, fruit bars or drinks, to prevent some chronic diseases where free radicals are involved. Also, El-Kafoury (2006) mentioned that Oreochromis niloticus received immunostimulant showed increase in albumen. globulin and total protein. In this regards, Abdel-Tawwab (2008) sated that Serum glucose, lipids, and protein, albumin, and globulin values of Oreochromis niloticus increased with the increase of Spirulina supplementation. Also in the same line with Kaoud et al, (2012) who added that the addition of dried Spirulina platensis to Oreochromis niloticus exposed to Hg improves the ALT and AST activity to be nearly as in the control. This study was conducted to determine the effects of graded levels of spirulina (A. plantensis) on some serum biochemical parameters and histopathological findings of Oreochromis niloticus.

Materials and Methods:

Fish: A total number of 270 O niloticus with an average body weight of 50±5 g were obtained from fish hatchery at the Central Laboratory for Aquaculture Research, Abbassa, Abo-Hammad, They Sharqia, Egypt. were transported in sterile plastic bags enriched containing water oxygen (2/3) to the lab of the Dept. of Fish Diseases, Faculty Veterinary Medicine, Suez Canal University. They were kept for two weeks under observation for acclimatization in glass aquaria (100×40×50cm). Fish were fed on the control diet for 2 weeks. The water was removed daily.

Aquaria: Fish were randomly distributed into six groups at a rate of 15 fish per aquaria. These aquaria were used for holding the experimental fish throughout the present period of the (triplicate each treatment). Each aguarium was supplied with chlorine free tap water (Innes, 1966) .The water temperature was kept at 22±1 °C. The continuous aeration was maintained in each aquarium using an electric pumping compressors.

Diet preparation: A basal diet was formulated to contain 30.6% crude protein diet table (1). The diet was daily provided at a fixed feeding ratio of 3% of body weight of fish according to *Eurell et al (1979)*. The daily amount of food was offered as two equal meals /day on two occasions over the day (9Am and12 PM).

S. platensis: used in the present study was obtained from Agent Chemical Laboratories. Redmond, WA, USA.

Blood sampling: At the 4 weeks and after 6 weeks, fish were fasted for 24 hours immediately prior to blood sampling and five fish per aquaria were randomly chosen. The blood was extracted from the caudal blood vessels. Blood was collected in eppendorf tubes with no anticoagulant in order to clot at 4°C

and centrifuged at 5000 rpm for 5 min at room temperature.

Serum biochemical examination: Glucose was determined calorimetrically according to Trinder (1969).Total content in serum was determined calorimetrically according to *Henry* (1964). Albumin and globulin in plasma were determined calorimetrically according to Drupt (1974). The serum level of AST and ALT determined was colorimeterically according to Reitman and Frankel (1957) serum creatinine level (mg/dl)estimated according to Henry et al (1974) serum uric acid (mg/d) was determined according to the method of Caraway (1963) and serum cholesterol (mg/d) was determined according to the method of Joseph et al (1972).

Challenge test: After month, fish from each treatment groups (10fish/

aquarium). Were challenged with pathogenic *Pseudomonas*. *fluorescens*. The fish in first group were injected intraperitoneally with 0.2 ml sterile saline containing (1.5 x 10° / ml) pathogenic strain, according to *El-Attar and Moustafa* (1996).

Histopathological examination: tissue specimen from liver and kidney were examined according to (*Drury and Willington*, 1980).

Statistical analysis: The obtained data were subjected to one-way ANOVA to evaluate the effect of spirulina supplement. Differences between means were tested at the 5% probability level using Duncan Multiple Range test. All the statistical analyses were done using SPSS program version 10 (SPSS, Richmond, VA, USA) as described by *Dytham* (1999).

Table 1: Design of the experiment of feeding of spirulina platensis to tilapia (O. niloticus):

Treatments (Groups)	Diet	Infection
I	Basal diet	Not infected
II	Basal diet containing 5g Spirulina/kg diet	Not infected
III	Basal diet containing 10g Spirulina/kg diet	Not infected
IV	Basal diet	Infected
V	Basal diet containing 5g Spirulina/kg diet	Infected
VI	Basal diet containing 10g Spirulina/kg diet	Infected

Results

Serum biochemical changes:

The results after 4 weeks showed that both transaminases enzymes (ALT and AST) were significantly decreased with the increase of spirulina supplementation compared to control. The results after 6 weeks showed that serum ALT and AST were stable in fish fed on spirulina while these enzymes were significantly increased in control group that fed on basal diet and infected with P. fluorescens group (IV).

The present study showed that fish fed on diets containing 10.0 g spirulina/kg group (III)diet exhibited higher total serum protein, albumin, and globulin as compared with fish fed the control diet (I). On the other hand the protein, albumin and globulin values were significant decreased in control group that fed on basal diet and infected with P. fluorescens, (group IV) significant increased values were in infected fish fed on obtained spirulina (groups V and VI) compared to control infected (group IV).

The glucose level was serum significantly increased in fish with increase of spirulina supplementation and the highest values were obtained at 1% spirulina group (group III) after4 weeks .While, serum glucose levels significantly increased in infected groups in which highest values obtained in 1% spirulina infected group (VI), the lowest values obtained in control group (IV) after 6 weeks.

The concentration of serum uric acid (mg/dl) and creatinine (mg/dl) of *O. niloticus* after 4 weeks were decreased by addition of spirulina in diet while, after 6 weeks showed that serum creatinine levels in non infected groups were decreased. Serum uric acid levels were stable in fish fed on spirulina. While the infected groups showed that the highest values of were obtained in control infected group (IV) and decreased by the increase of spirulina supplementation in diet.

After 4 weeks, serum cholesterol was significantly decreased in fish with the increase of spirulina supplementation. After 6 weeks serum cholesterol was increased in infected groups, the highest values in fish fed on basal diet infected with *P. fluorescens* (group IV), while cholesterol levels return to normal level by increase of spirulina supplementation in diet (table 2).

Histopathological results:

Histopathological results showed that hepatic melano-macrophage centers and Kupffer cells appeared more activated. Also, kidney and spleen showed activations of melano-macrophage centers incase of fish groups fed on spirulina supplemented feed than fish groups fed on basal diet.

However, the hepatic tissue of infected fish with P. fluorescens showed, In case of Group (IV): their liver showed congestion of

vein and hepatic hepatoportal sinusoid, hyperplasia of epithelial lining of bile duct, inactivation of pancreatic acini which infiltrated by leukcocyts and vacuolar hepatic degeneration of the parenchyma (figure 2).Group (V): hepatocyts showed mild swelling, hyperplasia of melanomacrophage center vacuolation and hepatocyts (figure 3). Group (VI): fish fed on basal diet and 1% infected spirulina and with pseudomonas fluorescence. their liver showed mild to moderate degree of degeneration (figure 4). Group IV, showed severe hydropic degeneration of the glomerlular and

renal tubular epithelium, variable depletion degree of of hemopiotic elements, congestion and focal hemorrhages of pertubular blood vessels (figure 6). Group V, showed mild congestion of some pertubular blood vessels, hyperplasia of melanomacrophage center and Hydropic degenerated of some renal tubules (figure Group VI, showed mild, focal degeneration of some tubular epithelium. The renal melanomacrophage centers were variably enlarged and hyper-activated and focal proliferation of the interstitial lymphoid elements also detected (figure 8).

Table (2) Serum biochemical changes of O. nloticus fed on practical diets containing different levels of spirulina after 4 weeks:

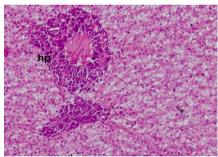
Commin	containing afferent levels of spiritina after 4 weeks.							
Items	Control (I)	Spirulina 0.5% (II)	Spirulina 1% (III)	Control infected (IV)	Spirulina 0.5% infected(V)	Spirulina 1% infected (VI)		
ALT (I.U./L)	13 ±0.20 a	12.7±0.15 a	7±0.23 b	13.2±0.69 a	12.8±0.15 a	6.9±0.51 b		
AST (I.U./L)	136±0.70 a	125±0.29 b	117±0.12 °	135.6±0.52 a	125.5±0.78 b	117.03±0.26 °		
Total protein (mg/dl)	3.4±0.35 b	3.6±0.26 ab	4.2±0.12 a	3.3±0.20 b	3.7±0.69 ab	4.1±0.69 a		
Albumin (mg/dl)	7±0.30b	2.2±0.12 b	2.73±0.88 ^a	2.1±0.06 b	2.23±0.15 b	2.6±0.34 ^a		
Globulin (mg/dl)	1.35±0.05 ^a	1.4±0.28a	1.5±0.03 ^a	1.33±0.03 ^a	1.44±0.28 ^a	1.86±0.21 ^a		
Glucose (mg/dl)	53.1±0.10	60±0.17 b	65±0.40 a	53.07±0.02 °	61.1±0.72 ab	65.4±0.42 a		
Creatinine (mg/dl)	0.39±0.01	0.2±0.02 b	0.14±0.03°	0.38±0.05 a	0.21±0.02 b	0.15±0.04 °		
Uric acid (mg/dl)	0.5±0.04 a	0.30±0.01 ^b	0.11±0.01°	0.49±0.03 ^a	0.32±0.02 ^b	0.16±0.05 °		
Cholesterol (mg/dl)	166±0.30 a	114±0.23 b	107±0.46 °	166.2±0.36 a	114.3±0.26 b	106.9±0.52 °		

Data in the same raw with different superscript are significantly different (P < 0.05)

Table (3) Serum biochemical changes of O. nloticus fed on practical diets containing different levels of spiruling after 6weeks:

containing different levels of spiritina after oweeks.							
Items	Control	Spirulina	Spirulina	Control	Spirulina0.5%	Spirulina 1%	
items	(I)	0.5% (II)	1% (III)	infected(IV)	infected (V)	infected (VI)	
ALT (I.U./L)	12.9±0.54 d	7±0.12 °	6±0.40 °	49±0.12 a	25±0.23 b	16±0.46 °	
AST (I.U./L)	135±0.05 ^d	118±0.17 e	105±0.35 ^f	235 ±0.23 a	201±0.36 b	181.3±0.44 °	
Total protein (mg/dl)	3.17±0.19 °	3.66±0.29 bc	5±0.29 ^a	2.9 ±0.26 °	3.3±0.12 bc	3.93±0.21 b	
Albumin (mg/dl)	2.2±0.12 ^{abc}	2.7±0.12ab	2.8±0.05 a	1.7±0.11 °	2±0.29 bc	2.17±0.39 abc	
Globulin (mg/dl)	0.96±0.49b	0.97±0.68 ^b	2.3±0.45 ^a	1.26±0.37ab	1.33±0.66 ^{ab}	1.55±0.21 ^{ab}	
Glucose (mg/dl)	65±0.60 ^f	82±0.30 °	106±0.70 b	96±0.40 d	100±0.70°	112±0.30 a	
Creatinine (mg/dl)	0.35±0.03 b	0.19±0.02 °	0.13±0.02 °	0.46±0.04 a	0.19±0.02 °	0.12±0.03 °	
Uric acid (mg/dl)	0.50±0.05 °	0.18±0.02 ^d	0.24±0.03 ^d	1±0.06 a	0.80±0.05 b	0.80±0.06 b	
Cholesterol (mg/dl)	150.6±0.40	132±0.23 ^d	137±0.12 °	173±0.23 a	150.9±0.32b	150.6±0.72 b	

Data in the same raw with different superscript are significantly different (P < 0.05)



Figure(1): liver of Tilapia fish, group control showing normal structure of hepatic cells and hepatopancreas (hp). H&E. X 400.

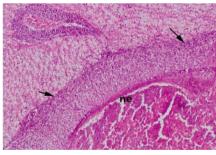


Figure (2): liver of Tilapia fish, group IV showing group massive necrosis of hepatic tissue (ne) & mononuclear cell infiltration (arrows). H&E. X 200.

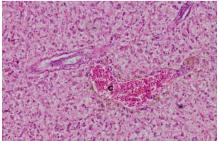


Figure (3): liver of Tilapia fish, group V, showing mild congestion of blood vessels (c). H&E. X 200.

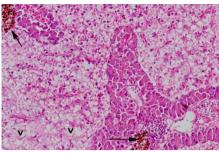


Figure (4): liver of Tilapia fish, group VI showing mild vacuolation (v) and activation of melanomacrophage centers (arrows). H&E. X 200.

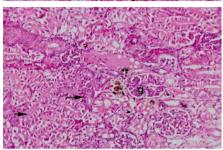


Figure (5): kidney of Tilapia fish, group I showing normal glomeruli (g) and renal tubules (arrows). H&E. X 200.

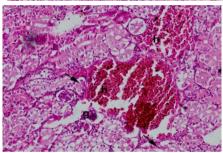


Figure (6): kidney of Tilapia fish, group IV showing hemorrhage(h), degeneration and necrosis of renal tubules (arrows).

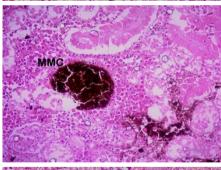


Figure (7): kidney of Tilapia fish, group V showing mild degeneration, activation of lymphoid tissue and activation of melanomacrophage centers (MMC). H&E. X 200.

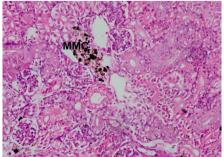


Figure (8): kidney of Tilapia fish, group VI showing activation of lymphoid tissue and melanomacrophage centers (MMC). H&E. X 200

Discussion

The activity of AST and ALT enzymes in blood may also be used as a stress indicator the significant changes in the activities of these enzymes in blood plasma indicate tissue impairment caused by stress (Svoboda, 2001). In the present study, enzymes (ALT and AST) were significantly increased in control group. This result agreed with Belal et al (2012), who revealed that fish fed with spirulina supplementation significantly decreased (AST) and (ALT) values compared with control. Also, in this concern, Ragap (2009) mentioned that the liver function enzymes serum S.GOT and S. GPT were stable in fish fed on spirulina while these enzymes were significantly increased in control group that feed in basal diet. The obtained results were proved by histological results hepatic where the melanomacrophage centers and Kupffer appeared activated cells more incase of fish groups fed on spirulina supplemented feed than fish groups fed on basal diet.

Also, the results after 6 weeks showed that serum ALT and AST were stable in fish fed on spirulina while these enzymes significantly increased in control groups. This agreed with Kaoud et al, 2012 who added that the addition of dried Spirulina platensis to *Oreochromis niloticus* exposed to Hg improves the ALT and AST activity to be nearly as in the control. In this regard, Wafaa

(2007) who revealed that increase ALT, AST liver enzymes in the experimentally infected fish with Pseudomonas fluorescens. The elevation in the ALT and AST attributed to the damage of hepatic cells and others tissues and escaping of these enzymes to the blood (Coles, 1986) and/or the increase in concentration of AST and ALT in blood plasma indicates impairment of parenchymatous organs mainly liver. In addition, the increase of plasma AST and ALT may be attributed to the hepatocellular damage or cellular degradation in liver, spleen or muscles (Yamawaki et al, 1986 and Kaoud et al, 2012). So indicating that spirulina has to some extent a protective role induced tissue injuries. The exact cause of this protective role in recovering tissue damages is not fully understood (Islam et al, 2009). Histopathological findings indicated that there were congestion of hepatoportal and hepatic vein sinusoid, hyperplasia of epithelial lining of bile duct. Inactivation of pancreatic acini which infiltrated by leukcocyts vacuolar and degeneration of hepatic the parenchyma of fish group fed on normal diet and infected with pseudomonas fluorescence (control +v) While, groups V and VI showed regeneration (mild to moderate regenerations) with hyperactivity of melenomacrophage centers of haemopiotic organs. This results may be due to the effect of extracellular proteases produced by

P. fluorscens which attack endothelial lining of blood vessels and parenchymatous organ causing the haemorrhagic phenomena as well as the degenerative changes (Ehab, 1991).

In the present study, the results showed that fish fed on diets 10.0 g Spirulina/kg containing group (III) diet exhibited higher total protein, albumin, and globulin as compared with fish fed the control diet (I). These results were parallel to that obtained by Abdel-Tawwab et al (2008), Ragap(2009) , Andrews et al (2011) and Belal et al (2012) who revealed that the serum total protein, albumin, and globulin values were increased by addition of spirulina in Commonly, increases in the levels of plasma total protein, albumin and globulin in fish are thought to be associated with a stronger innate immune response (Wiegertjes et al, 1996). On the other hand the albumin protein, and globulin values were significant decreased in control group that fed on basal diet and infected with P. fluorescens, while significant (group) IV) increased values in infected fish fed on spirulina(groups V and VI). In this regards, Sivagurunathan et al (2012) recorded that significant increase was observed in serum total protein and albumin levels of Cirrhinus mrigala infected with P. fluorescens fed on Lotus for 40 days. The increase in serum protein content might be in part due to an increase in the WBC, which is a major source of serum protein production such as lysozyme, complement factors and bactericidal peptides (*Misra et al, 2006*).

The blood glucose level is a sensitive indicator in fish environmental stress (Silbergeld, 1974). The result of this study after 4 weeks showed that the glucose level was significantly increased in fish with the increase of spirulina supplementation. These results were parallel to that obtained by Abdel-Tawwab et al (2008) and Belal et al (2012), that fish fed with diets containing 1% spirulina exhibited higher glucose levels. These results may be attributed to its bioactive components such as 13.5% carbohydrates; the sugar composition is mainly composed of along with rhamnose, glucose, mannose, xylose, galactose, and two 2-O-mehtyl-lsugars: unusual 3-Omethyl-lrhamnose and rhamnose. (Madhava et al, 2000). Also, after 6 weeks showed that serum glucose levels were significantly increased in infected groups. The elevation in glucose level due to stress and stimulation of glycogenolysis (Soivio et al. 1974). And/or may be due to mobilization of glucose in response to stress is generally accepted as a mean of providing extra energy resources enabling the fish to overcome the disturbance (Arends et al, 1999).

The concentration of uric acid (mg/dl) and serum creatinine (mg/dl) of *O. niloticus* after 4 weeks

were highest in control group fed on basal diet (I). This result clearly shows the protective effect of spirulina that may be due to Cphycocyanin which is one of the major constituent of spirulina protect renal cell injury (Faroog et al, 2004). After 6weeks the infected groups showed that the highest values were obtained in control infected group (IV) and decreased by the increase of spirulina). The elevation in serum creatinine and serum uric acid levels may be due to alteration in normal physiology of kidney due to bacterial toxin. (Coles, 1986).

After 4 weeks serum cholesterol was significantly decreased in fish with the increase of spirulina. While. after 6 weeks serum cholesterol was increased in which the infected groups, in highest values were obtained in control infected group. results agreed with Heidarpour et al (2011) who revealed that the fish (Holstein calves) showed significant reduction in plasma cholesterol by increasing spirulina in diet. This cholesterol serum reduction has been stated as the effect of spirulina on lipoproteins metabolism and the increase of the lipoprotein enzyme activity levels (Karkos et al. 2008). The obtained results were proved by histological results where hydropic degeneration of the glomerlular and renal tubular epithelium, variable depletion degree of of the hemopiotic elements, congestion and focal hemorrhages of the

pertubular blood vessels of fish group fed on normal diet and infected with pseudomonas fluorescence (control +v) While. VI showed groups V and regeneration (mild to moderate regenerations) with hyperactivity of melenomacrophage centers of haemopiotic organs.

The present study concluded that Spirulina positively improved serum biochemical parameters of Oreochromis niloticus, as well as its challenge resistance to by infections. fluorescens Τt recommended supplement to spirulina in the diet of Nile tilapia especially those grow in farms under immunosuppressive/stressful conditions.

References

Abdel-Tawwab, M.; Ahmad, M. H.; Abdel-Hadi, Y. M. and Seden, M. E.A. (2008): Use of spirulina (Arthrospira platensis) as agrowth and immunity promoter for Nile tilapia, Oreochromis niloticus (L) fry challenged with pathogenic Aeromonas hydrophila. International symposium on tilapia in aquaculture, 1015-103.

Andrews, S. R.; Sahu, N. B.; Pal, A. B.; Mukherjee, S. C. and Kumar, S. (2011): Yeast extract, brewer's yeast and spirulina in diets for Labeo rohita fingerlings affect haemato-immunological responses and survival following Aeromonas hydrophila challenge. Research in Veterinary Science, (91):103–109.

Arends, R. J.; Mancera, J. M.; Munoz, J. L/ and Wendelaar Bonga, S. E. (1999): The stress response of the gilthead sea bream (Sparus aurata L.) to air exposure and confinement J. Endocrin., 163: 149-157.

Belal, E. B., Khalafalla, M. M. E. and El-Hais, A. M. A. (2012): Use of spirulina (Arthrospira fusiformis) for promoting growth of Nile Tilapia fingerlings. Afr. J. Microbiol. Res (35), 64

Bermejo, P.; Piñero, E. and MaVillar, A. (2008): Iron-chelating ability and antioxidant properties of phycocyanin isolated from a protean extract of Spirulina platensis. Food Chemistry, 110: 436–445.

Caraway, W. T. (1963): Standard Method of Clinical Chemistry. Academic press,

Coles E, H. (1986): Text book of veterinary clinical pathology. 4 th ed. W.B. Sounders Co., Philadelphia, London. Drupt, F. (1974): Colorimetric method for determination of albumen. Pharm. Bio., 9: 777-779.

Drury, R.; Wallington, E. and Cancerson, R. (1976): Carleton's histological technique". 4 th ed. Oxford University Press, London.

Dytham, *C.* (1999): Choosing and Using Statistics: A Biologist's Guide. Blackwell Science Ltd., London, UK.

Ehab El-Din, A. E. F. (1991): Effect of some stresses on pseudomonas fluorescens infection in fresh water fishes. M. V. Sc.,

thesis, Faculty of Vet. Mid. Cairo University.

El-Attar, M. and Moustafa, A. (1996): Experimentally infected Tilapia fish (I/P) with 0.5 ml broth cultured (3x107 cell /ml) of *P. fluorescens.* Assiut. Vet. Med. 35: 155-162.

El-Kafoury, *M*. \boldsymbol{A} . (2006): Comparative studies between **Oreochromis** niloticus and Monosex tilapia from immunological and pathological aspect of view.MVSc.(Dep.fish disease and management) Of Vet. Med. Alex. Univ.

Eurell, T.; Lewis, S. and Grumbles, L. (1979): Comparison of selected diagnostic tests for detection of motile Aeromonas septicaemia in fish. Am. J. Vol. Res. 1979; 39 (8): 1384 – 1386.

Farooq, S.M.; Asokan, D.; Sakthivel, R.; Kalaiselvi, P.; Varalakshmi, P.; (2004): Salubrious effect of C-phycocyanin against oxalate mediated renal cell injury. Clin Chim Acta (348),199–205.

Heidarpour, A.; Shahraki, A. D. F. and Saied, S. H. (2011): Effects of Spirulina platensis on performance, digestibility and serum biochemical parameters of Holstein calves." African Journal of Agricultural Research 6 (22), pp. 5061-5065

Henry, R. J. (1964): Colorimetric determination of total protein In: Clinical Chemistry. Harper and Row Publ., New York, USA, p 181.

Henry, R. J.; Harper, R. and Hagerstion, R. (1974): Clinical

Chemistry, Principles and Technique. 2nd ed. HP. Co. Philadelphia.

Ibrahem. M. D., Fathi, M., Mesalhy, S., & Abd El-Aty, A. M. dietary (2010): Effect of supplementation of inulin and vitamin C on the growth. hematology, innate immunity, and resistance of Nile tilapia (Oreochromis niloticus). Fish Shellfish Immunol, 29, 241-6.

Innes, *W. T.* (1966): Exophic Aquarium fishes. 4th ed.Aquc. Inc. Jersy, USA.530-533.

Islam, M. S.; Awal, M.; AMostofa, M.; Khair, F. A. and Myenuddin, M. (2009): Effect of Spirulina on Biochemical Parameters and Reduction of Tissue Arsenic Concentration in Arsenic Induced Toxicities in Ducks. International Journal of Poultry Science 8 (1): 69-74.

Joseph, A.; Knight, M.; Anderson, S.; James, M. and Rawie, H. (1972): Chemical basis of the sulfophospho-vanillin reaction for estimating total serum lipid. Cl. Chem., 18(3), 198-201.

Kaoud, H. A.; Mahran, K. H. A.; Rezk, A. and Khalf, M. A. (2012): Bioremediation the toxic effect of mercury on liver histopathology, some hematological parameters and enzymatic activity in Nile tilapia, Oreochromis niloticus. Researcher; 4(1), 60-70.

Karkos, P.D.; Leong, S.C.; Karkos, C.D.; Siraji, N.; Assimkapoulos, D.A. (2008): Review of spirulina in clinical practice: Evidence-Based

human applications. e CAM Adv. Access, (14), 1-4.

Lin, W.; Pan, B.; Sheng, J.; Xu, J. and Hu, Q. (2007): Antioxidant activity of Spirulina platensis extracts by supercritical carbon dioxide extraction. Food Chem., (105), 36–41.

Madhava, C.; Bhat, V. B.; Kiranmai, G.; Reddy, M. N.; Reddanna, P. and Madyastha, K. M. (2000): Selective inhibition of cyclooxygenase-2 by C-phycocyanin, a biliprotein from Spirulina platensis. Bioch. Biophy. Res. Comm., (277), 599–603.

Misra, C. K.; Das, B.K.; Mukherjee, S.C. and Meher, P.K. (2006): The immunomodulatory effects of tuftsin on the non-specific immune sstem of Indian Major Carp. Labeo *rohita*. Fish shellfish Immunol(20),728-738.

Ragap H M. (2009): Some studies on Spirulina on some cultured freshwater fish in Egypt. MVSc. (Dep.fish disease and management) Vet. Med. Alex.Univ.

Reitman, S. and Frankel, S. (1957): Colorimetric determination of glutamic oxaloacetic and glutamic pyruvic transaminases. Am. J. Clin. Pathol. (28).

Silbergeld, E. K. (1974): Blood glucose: a sensitive indicator of environmental stress in fish. Bull. Envir. Contam. Toxic, 11: 20-25.

Sivagurunathan, A.; Innocent, X. B. and lakshmi, S. M. (2012): Immunomodulatory Effect of Dietary Nelumbo Nucifera (Lotus) in Growth and Haematology of

Cirrhinus Mrigala Challenged With Pseudomonas Cirrhinus Mrigala Challenged With Pseudomonas. Journal of Applied Pharmaceutical Science 02 (07), 191-195.

Soivio A., Westman K. and Nyholm K. (1974): The influence of changes in oxygen tension on the haematocrit value of blood samples from asphyxic rain bow trout (salmo gairderi) .Aquaculture, 3: 395-401.

Svoboda, M. (2001): Stress in fish – review. Bul. VURH Vodnany, (37), 69-191.

Trinder, *P.* (1969): Determination of glucose concentration in the blood. Ann.Clin.Biochem, 6-24.

Wafaa, *Z*. *A*. (2007): Clinicopathological studies on the

effect of pseudomonas infection in fresh, water fish. PhD. Thesis Fac. of Vet.(Clinical Pathologhy) Med. Suez canal.Univ.

Wiegertjes, G. F.; Stet, R. J.; Parmentier. Н. *K*.: Van W. В. Muiswinke. (1996): **Immunogenetics** of disease resistance in fish; a comparable approachDev. Comp. Immunol. (20), 365-381.

Yamawaki, K.; Hashimoto, W.; Fujii, K.; Koyama, J.; Ikeda, Y. and Ozaki, H. (1986): Hematological changes in carp exposed to low cadmium concentration. Bull of the Japanese. Soc. Sci. Fish., 59 (3):459-466.

الملخص العربي

بعض الدراسات الباثولوجيا الإكلينيكية على تأثير السبيرولينا فى سمك البلطى النيلى أسامة على محمد، أسماعيل عبدالمنعم عيسى،أمنية السيدكيلاني، امينة على دسوقى شيماء محمد البحار

مازال إستخدام الطحالب الدقيقة في مجال تغنية الأسماك يتم على نطاق ضيق. وقد أجريت هذه الدراسة بهدف دراسة التاثيرات الناتجة عن إضافة طحلب السبيرولينا (السبيرولينا بلاتنسيس) لأعلاف أسماك البلطي النيلي بمستويات مختلفة من مسحوق الطحالب. وقد إستخدم في هذه الدراسة ١٧٠سمكة بلطي نيلي تم تجميعه من مزرعة مركز بحوث الأسماك بالعباسة بأبوحماد شرقية وكان متوسط وزن السمكة الواحدة ٥٠ جرام تقريبا وقد أحضرت الأسماك إلى المعمل بقسم أمراض ورعاية الأسماك بكلية الطب البيطري جامعة قناة السويس في أكياس بلاستيكية مزوده بنسبة أكسجين ١٦٠٣. تم إستخدام ٣ علائق تجريبية بإضافة مسحوق الطحالب في علائق الأسماك المتزنة بنسب مختلفة وقد تم تقسيم الأسماك إلى ست مجموعات بحيث كل مجموعة أسماك مقسمة إلى ثلاث مكررات لكل معامله لكل معامله يحتوي كل مكرر على عدد ١٥ سمكة في أحواض زجاجيه وذلك لمدة ستة اسابيع وتركت هذه الأسماك للأقلمة لمدة أسبوعين قبل بداية التجربة.

- المجموعة الأولى: تم تغذيتها على العليقة الأولى والتي تتكون من عليقة أسماك متزنة
- المجموعة الثانية: تم تغذيتها على العليقة الثانية والتي تتكون من عليقة أسماك متزنة مضاف إليها نسبة ٥,٠% من مسحوق نفس الطحلب.

• المجموعة الثالثة: - تم تغذيتها على العليقة الثالثة والتي تتكون من عليقة أسماك متزنة مضاف المجموعة الثالثة الشهدد المجموعة المتالثة المتحدد المتحدد

- المجموعة الرابعة: تم تغذيتها على العليقة الأولى والتي تتكون من عليقة أسماك متزنة بدون إضافة مسحوق نفس الطحلب لها وعرضت للعدوى بميكروب السودو موناس فلورسنس في الغشاء البريتوني بجرعة ٢٠,٠مل في نهاية التجربة.
- المجموعة الخامسة: تم تغذيتها على العليقة الثانية والتى تتكون من عليقة أسماك متزنة مضاف إليها نسبة ٥٠٠% من مسحوق نفس الطحلب وعرضت للعدوى ببنفس الميكروب في نهاية التجربة.
- المجموعة السادسة: تم تغذيتها على العليقة الثالثة والتي تتكون من عليقة أسماك متزنة مضاف البيها نسبة ١% من مسحوق نفس الطحلب وعرضت للعدوى ببنفس الميكروب في نهاية التجربة. وأسفرت هذه الدراسة عن النتائج التالية:أوضحت الدراسة زيادة في نسبة الألبيومين الجاوبيولين الماليروتين الكلى في مجاميع الأسماك التي تغذيت على عليقه مزودة بمسحوق طحلب السبيرولينا بالمقارنة بالمجموعة الأولى. وقد لوحظ انخفاض في مستوى أنزيمات الكبد في مجموعات الأسماك المغذاه على علي علي علي علية أسماك مذودة بمسحوق طحلب السبيروليينا في جميع المعاملات خلال فترة التجربة بالمقارنة بالمجموعة الأولى. بعد العدوى بميكروب السودوموناس لوحظ زيادة بأنزيمات الكبد في المجموعة الأولى مع ثبات الأنزيمات للمعدل الطبيعي في المجموعة الخامسة والسادسة. كما كان هناك ارتفاعا واضحا في نسبة الجلوكوز في المجموعة الخامسة والسادسة. كما كان هناك ارتفاعا واضحا في نسبة الجلوكوز في المجموعة الخامية المجموعات المغذاة في نسبة الكوليسترول في المجموعات المغذاة في جميع المجموعات وحد ايضا انخفاضا ملحوظا في نسبة الكوليسترول في المجموعات المغذاة في جميع المجموعات المغذاة المعدل المعدل المعدل المعدا المعدوعات المغذاة في نسبة المجموعات المغذاة في نسبة الكوليسترول في المجموعات المغذاة في جميع المجموعات المغذاة المعدا المعدا

ومع إمتداد فترة التجربة بالمقارنة بالمجموعة الأولى وجد ان هناك زيادة في نشاط خلايا الميلانومكروفاج الموجودة في الكبد والطحال والكلى بالأسماك المغذاة على العليقة المضاف إليها السبيرولينا. اما بعد العدوى فكانت هناك تغييرات هدامة في خلايا الكبد مع ارتشاح خلوى في المجموعة الرابعة، اما المجموعة السادسة فكان هناك زيادة في نشاط خلايا الميلانوميكروفاج.

على السبير ولينا